Western North American Naturalist 68(1), © 2008, pp. 83-102

GEOGRAPHIC PATTERNS OF GROUND-DWELLING ARTHROPODS ACROSS AN ECOREGIONAL TRANSITION IN THE NORTH AMERICAN SOUTHWEST

David C. Lightfoot^{1,3}, Sandra L. Brantley¹, and Craig D. Allen²

ABSTRACT.—We examined the biogeographic patterns of ground-dwelling arthropod communities across a heterogeneous semiarid region of the Southern Rio Grande Rift Valley of New Mexico. Our 3 sites included portions of 5 ecoregions, with the middle site a transition area where all ecoregions converged. We addressed the following 3 questions: (1) Do the species assemblage patterns for ground arthropods across habitats and sites conform to recognized ecoregions? (2) Are arthropod assemblages in distinct vegetation-defined habitats within an ecoregion more similar to each other or to assemblages in similar vegetation-defined habitats in other ecoregions? (3) Is there a detectable edge effect with increased arthropod diversity in the area of converging ecoregions? We encountered 442 target arthropod species from pitfall traps operating continuously for 7 years over a series of different habitats at each of the 3 sites. We examined geographic distributions of spider and cricket/grasshopper species in detail, and they showed affinities for different ecoregions, respectively. Each habitat within a study site supported a unique overall arthropod assemblage; nevertheless, different habitats at the same site were more similar to each other than they were to similar habitats at other sites. Overall arthropod species richness was greatest in the area where all 5 ecoregions converged. Arthropod species and their geographic distributions are poorly known relative to vascular plants and vertebrate animals. Findings from this research indicate that ecoregional classification is a useful tool for understanding biogeographic patterns among arthropods.

Key words: biome transition, crickets, diversity, ecoregion, ecotone, edge effect, grasshoppers, insects, species, spiders.

Geographic distribution patterns of plant and animal species result from multiple factors, including proximity to historical and evolutionary source areas, climate, distribution and availability of environmental resources, and biotic interactions (Ricklefs and Schluter 1993, Brown and Lomolino 1998, Lomolino and Heaney 2004). Much is known about the geographic distribution and diversity patterns of vascular plants and vertebrate animals, however, relatively little is known about the geographic patterns and processes accounting for the distributions of most terrestrial invertebrates (Parmenter et al. 1995, Ricketts et al. 1999). One of the goals of our research was to provide much-needed information on the distribution and abundance patterns of terrestrial arthropods.

At the continental scale, conceptual biomes (Whittaker 1975, Ricklefs and Miller 2000), or more finely delineated ecoregions (e.g., Ricketts et al. 1999), are biogeographic areas with similar climates, landscapes, and biotic assemblages that are more similar to each other than to those in adjacent ecoregions. The association of species with biogeographic regions includes abiotic factors (especially climate) and biotic factors such as plant-animal and predator-prey interactions (Shmida and Wilson 1985, Brown 1995). Species richness is expected to increase at ecotones (edge effect of Odum 1971, Holland et al. 1991, Risser 1993, 1995), the areas where different regional assemblages converge. The extent of species overlap depends on individual species' distributions, the number and steepness of environmental gradients, and the number of converging biogeographic regions.

The American Southwest is geologically young, with a flora and fauna that developed mostly during the Quaternary Period (Axelrod 1958, Van Devender and Spaulding 1983, Van Devender 1995). The area supports some of the most topographically (Hunt 1983) and biologically diverse semiarid temperate landscapes in the world (Brown 1982, Parmenter et al. 1995), ranging from hot, dry lowland deserts to cool, moist montane forests. Within the Southwest, several continental ecoregions (Ricketts et al. 1999) converge across the Southern Rio Grande Rift Valley that bisects New Mexico from north

¹Museum of Southwestern Biology, University of New Mexico, Albuquerque, NM 87131.

²United States Geological Survey, Biological Resources Discipline, Jemez Mountains Field Station, Los Alamos, NM 87544. ³E-mail: dlightfo@unm.edu

s-man: ungnuo@unini.euu

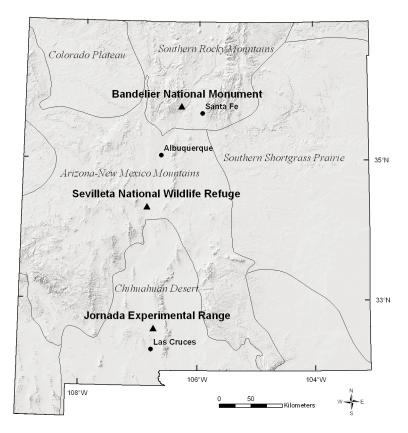


Fig. 1. Map of the study area and ecoregion boundaries.

to south (Hawley 1978): forests of the Southern Rocky Mountains (sensu current Nature Conservancy / World Wildlife Fund designation) and forests of the Arizona-New Mexico Mountains (sensu current Nature Conservancy / World Wildlife Fund designation) occupy the high elevations; and the Colorado Plateau, the Chihuahuan Desert, and the Western Short Grasslands (Southern Shortgrass Prairie, sensu current Nature Conservancy / World Wildlife Fund designation) all converge in the lowlands (Fig. 1). Additionally, the Southern Rio Grande Rift Valley spans the convergence of 3 geographically higher-ranking bioregions: eastern North America to the east, northern Mexico to the south, and western North America to the north (Ricketts et al. 1999).

The goal of this study was to assess geographic patterns of arthropod communities associated with landscapes across this area of ecoregion transitions. We focused on grounddwelling arthropods, which have important roles in semiarid systems as detritivores, herbivores, and predators (Crawford 1981, Whitford 1986, Polis 1991). To better understand the biogeography of these arthropods, we addressed the following questions: (1) What are the species assemblage patterns for ground arthropods across the study sites and across within-site habitats, and do they conform to recognized ecoregions (sensu Ricketts et al. 1999)? (2) Are arthropod assemblages in distinct vegetation-defined habitats within an ecoregion more similar to each other than to assemblages in similar vegetation-defined habitats in other ecoregions? (3) Are there detectable edge effects (Odum 1971) for increased arthropod diversity in the transition area of converging ecoregions?

Methods

Study Sites

This study was conducted at 3 sites located along the Southern Rio Grande Rift Valley in New Mexico (Fig. 1). A series of 3–4 local

84

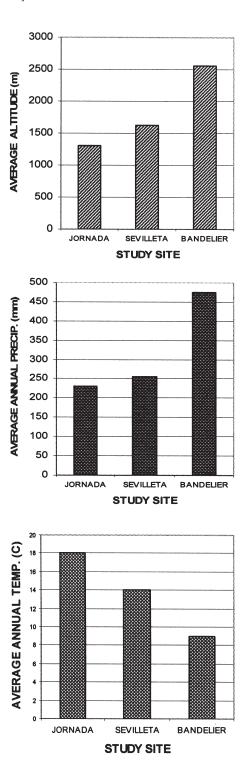


Fig. 2. Average elevations (m asl) (a), long-term average annual precipitation (b); and long-term average daily ambient temperatures (c) from each of the 3 principal study sites.

habitats recognized on the basis of plant community classifications (Dick-Peddie 1993) were sampled within each study site, for a total of 10 habitat samples across a biogeographic transition region from the Chihuahuan Desert to the south, the Southern Rocky Mountains to the north, the Arizona–New Mexico Mountains and Colorado Plateau to the west, and the Southern Shortgrass Prairie to the east (Ricketts et al. 1999; Fig. 1). Dick-Peddie (1993) and Brown (1982) provide detailed hierarchical vegetation classifications for the floristic and environmental characteristics of this region.

The 3 study sites were associated with 3 different long-term ecological research programs: the National Science Foundation (NSF)– funded Jornada Basin Long-Term Ecological Research (LTER) program, the NSF Sevilleta LTER program, and the United States Geological Survey, Jemez Mountains Field Station. The habitats used for this study were representative of the ecoregions listed above and were the dominant environments across each of the 3 research sites.

The Chihuahuan Desert ecoregion (Ricketts et al. 1999), consisting of Chihuahuan Desert scrub and desert grassland communities (Dick-Peddie 1993) in the southern Rio Grande valley of New Mexico, was represented by 4 habitats in the Jornada Basin (JRN) in south central New Mexico: (1) mesquite (*Prosopis gland-ulosa*) sand dune fields, 1330 m, 32°33'0"N, 106°42'39"W (datum NAD27 for all coordinates); (2) tarbush (*Flourensia cernua*) basin floor, 1315 m, 32°37'09"N, 106°47'13"W; (3) creosotebush (*Larrea tridentata*) piedmont slopes, 1355 m, 32°40'04"N, 106°46'12"W; and (4) black grama (*Bouteloua eriopoda*) grassland, 1318 m, 32°29'22"N, 106°46'47"W.

The Sevilleta National Wildlife Refuge (SEV) in the central Rio Grande valley of New Mexico was geographically centered between the JRN to the south and Bandelier (see below) to the north. The SEV also represented a region of multiple ecoregional transitions from the north, south, east, and west. The SEV was located within the Arizona–New Mexico Mountains ecoregion, yet the lowland environments contained flora characteristic of the surrounding low-elevation Chihuahuan Desert and the Southern Shortgrass Prairie ecoregions. We sampled 3 principal habitats at the SEV: (1) creosotebush piedmont (Chihuahuan Desert ecoregion; Ricketts et al. 1999), 1615 m,

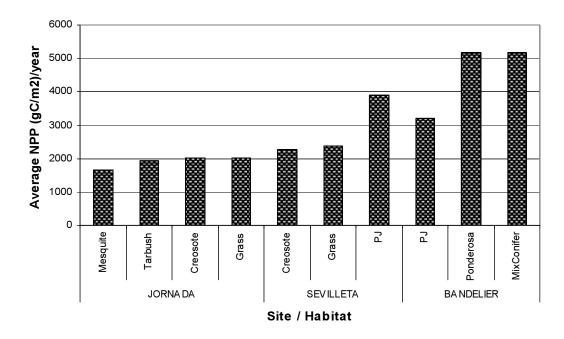


Fig. 3. Average annual net primary production among study sites and habitats estimated from MODIS remote sensing data, averaged over the years 2000–2002.

34°19′59.0″N, 106°44′9″W; (2) black grama grassland (Chihuahuan Desert ecoregion) mixed with some blue grama (*Bouteloua gracilis*; Southern Shortgrass Prairie and Colorado Plateau), 1616 m, 34°20′17.0″N, 106°43′3″W; and (3) pinyon-juniper (*Pinus edulis* and *Juniperus monosperma*, respectively) woodland (Arizona–New Mexico Mountains forest ecoregion), 1976 m, 34°22′6.3″N, 106°32′6″W.

Three habitats at Bandelier National Monument (BNM) in the Jemez Mountains at the north end of the Southern Rio Grande Rift valley represented higher-elevation ecoregions and formed an elevational gradient: (1) pinyonjuniper woodland (Arizona-New Mexico Mountains forest ecoregion), 1948 m, 35°49'47"N, $106^{\circ}22'4''W$, with variable densities of pinyon (P. edulis) and juniper trees (J. monosperma); (2) ponderosa pine (Pinus ponderosa) forest (Arizona-New Mexico Mountains forest ecoregion). 2454 m, 35°45′45″N, 106°15′52″W, with moderate to open conifer canopies; and (3) montane mixed conifer forest (P. ponderosa, Pseudotsuga menziesii, Pinus strobiformis, Abies concolor, Populus tremuloides; Southern Rocky Mountains forest ecoregion), 2712 m, 35°51'6"N, 106°24′41″W, with dense conifer canopy.

Elevation and long-term average precipitation increased from JRN to BNM (Fig. 2a–b), but long-term average annual ambient temperatures decreased (Fig. 2c). We used moderate resolution imaging spectroradiometer (MODIS) remotely-sensed data (Zhao et al. 2005, Zhao and Running 2006) averaged from 2000 to 2002 to estimate aboveground plant net primary production (ANPP; a 1-km pixel centered on pitfall traps within each habitat type per site). Plant productivity at JRN was consistently lower than at SEV or BNM; estimated ANPP values were highest at BNM (Fig. 3)

Arthropods

Ground arthropods were sampled from pitfall traps (Southwood 1966, Digweed et al. 1995, Ward et al. 2001, Woodcock 2005) consisting of small plastic cups (7.5 cm top diameter \times 10 cm deep, 0.25-L volume) placed inside small steel cans (7.5 cm top diameter \times 10 cm deep) that were dug into the ground until the top of the can was level with the soil surface. Propylene glycol was used as the preservative, and tile covers (or local rocks at BNM where elk [Cervus elaphus] were attracted to artificial covers) protected the samples from weather. Replicate lines of traps (3 at JRN, 5 at SEV, and 5 at BNM) 30-50 m long were placed in each habitat type at each study site. Arthropod species richness and arthropod

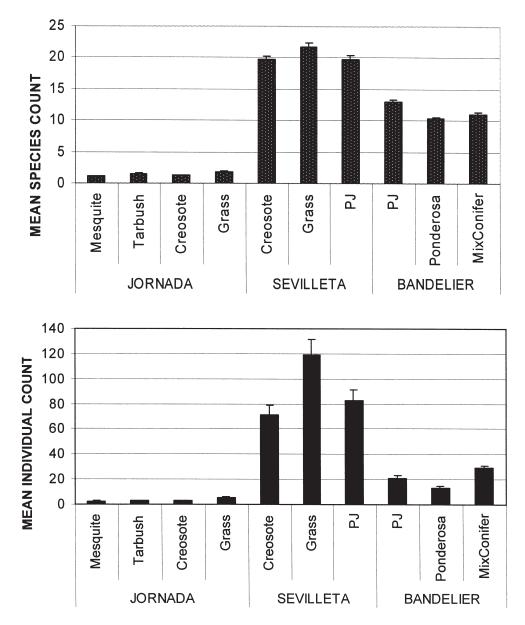


Fig. 4. Ground arthropod species richness among different habitats across the 3 principal study sites (a). Ground arthropod abundance across different habitats across the 3 principal study sites (b). Values are means per trap per replicate plot per year, with 1 standard error, over a 7-year period (1996–2002).

abundance were averaged over all traps by habitat. Trap line locations were subjectively chosen to represent a range of environments within each habitat type. Traps were left open continuously and arthropods were collected every 2 months from 1996 to 2002. Trap lines within habitats were located within 10 km of each other, and all habitat types within a study site were no more than 20 km apart. Target taxa (i.e., taxa that we studied) included only those arthropods that are known to live primarily on the ground and that are appropriately sampled by pitfall traps (Uetz and Unzicker 1976, Thomas and Sleeper 1977, Adis 1979): spiders (4 families), bristletails (1 family), crickets (3 families), grasshoppers (2 families), true bugs (2 families), and beetles (4 families), as well as scorpions, sun spiders, centipedes, and

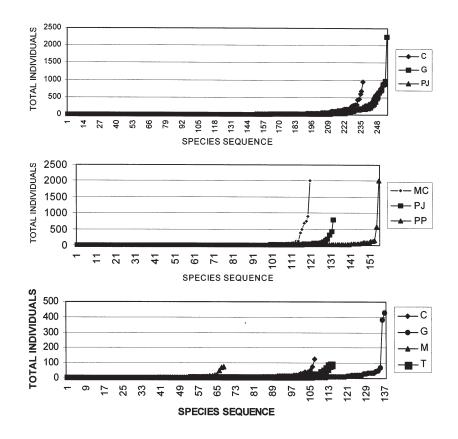


Fig. 5. Species richness and abundance curves for ground arthropods from the Sevilleta (a), Bandelier (b), and Jornada (c) habitats from 1996 to 2002.

millipedes (see Appendix 1 for scientific names of classes, orders, and families). Taxonomic classification followed Arnett (2000) for ranks from the class to the subfamily level and Poole and Gentili (2003) for genera and species. We used published distribution information to analyze patterns of grasshoppers/crickets and spiders in detail (Appendix 2). Two of us were specialists on the taxonomy, biogeography, and ecology of grasshoppers (D.C. Lightfoot) and spiders (S.L. Brantley) relative to other arthropod groups. Voucher specimens representing all target taxa were deposited in the Museum of Southwestern Biology, Division of Arthropods, University of New Mexico, Albuquerque.

Data Analysis

Multivariate and univariate data analyses were performed with PC SAS (Statistical Analysis Systems, v. 8.2; SAS Institute, Inc. 2003) software. We used nonparametric agglomerative hierarchical unweighted pair-group average cluster analysis (SAS, PROC CLUS) as a descriptive method to examine similarities of arthropod species assemblages among and within habitats, based on Euclidean distance as a measure of the similarity of species composition between habitats and sites. Only species that were represented by 10 or more observations over the entire study period were included in that analysis, thereby emphasizing only relatively common or important taxa, not rare species that may not have been adequately sampled. We used log-transformed count data for canonical discriminant function analysis (CDFA; SAS, PROC CANDISC) to test the hypothesis of no spatial differences among site habitats based on arthropod taxa/ trophic groups (i.e., the family-level ranks listed above), and to provide ordinations of habitats within sites based on arthropod composition. Large numbers of species encountered and unequal sample variances associated with various species counts (see results section, Fig. 4a-c) precluded us from using CDFA with species-level data.

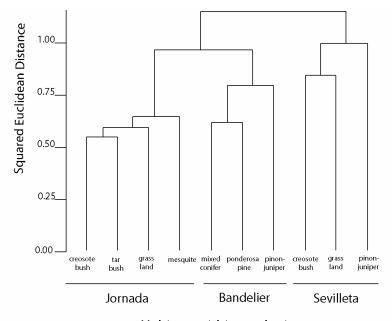




Fig. 6. Cluster analysis dendrogram of the different habitats from the 3 study sites based on ground arthropod species composition. Group average linkage method scaled by squared Euclidean distance. Habitat codes: con = mixed conifer, cre = creosotebush, gra = grassland, mes = mesquite, pnj = pinyon-juniper, pon = ponderosa pine.

RESULTS

Species Diversity Patterns of Ground Arthropod Assemblages

We encountered a total of 442 target taxa across the 3 study sites over the 7-year period (Appendix 1). The greatest species richness was at SEV with 306 species, followed by BNM with 190 species and JRN with 144 species.

The greatest species richness was at the SEV black grama grassland followed by the SEV pinyon-juniper woodland and creosotebush habitats (Fig. 4a). The SEV grassland had the highest species richness and highest overall abundance of individuals (Fig. 5a). Species richness among BNM montane habitats was lowest at the high-elevation montane mixed conifer and midelevation ponderosa pine forests, and greatest at the low-elevation pinyon-juniper woodland habitat (Figs. 4a, 5b). Species richness across the 4 JRN Chihuahuan Desert habitats was greatest in the black grama grassland, followed by the tarbush, creosotebush, and mesquite sand dune habitats, respectively (Figs. 4a, 5c).

Abundance or counts of arthropod taxa followed patterns similar to those of species richness (Figs. 4b, 5). Ground arthropod counts averaged over the 7-year study period were greatest at the SEV sites, of which the black grama grassland habitat supported by far the greatest numbers of arthropods (Fig. 4b). BNM had the 2nd-greatest overall arthropod abundance, ranging from highest in the montane conifer forest, followed by the pinyon-juniper woodland and then the ponderosa pine forest. The Chihuahuan Desert habitats at JRN had the lowest overall ground arthropod counts, with greatest abundance within the black grama grassland.

Spatial Patterns of Ground Arthropod Habitat Associations

Cluster analysis of all 10 habitats from the 3 study sites based on ground arthropod species composition resulted in squared Euclidean distances >0.50 among all habitats and a distance of about 1.00 among the 3 sites (Fig. 6). The JRN creosotebush and tarbush habitats were most similar in ground arthropod species composition, followed by grassland and mesquite sand dune habitats. At BNM, montane conifer and ponderosa pine forests were more similar to each other than to the pinyon-juniper

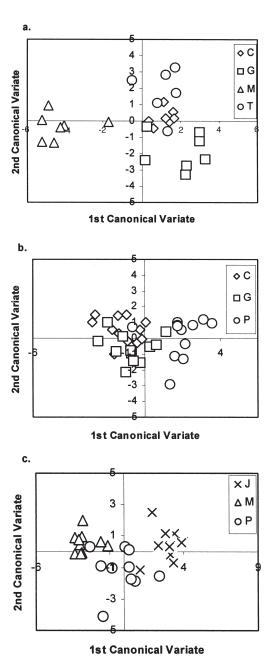


Fig. 7. Canonical discriminant function analysis ordination of Jornada (a), Sevilleta (b), and Bandelier (c) arthropod assemblages samples from the different habitats based on ground arthropod family-level composition and abundance averaged over a 6-year period. See text for explanation of canonical variates.

woodland, and at SEV, creosotebush and black grama grassland habitats were more similar to each other than to the pinyon-juniper woodland. Arthropod species assemblages among different habitats within each of the 3 study sites were more similar to each other than to similar habitats across the 3 study sites: black grama grassland and creosotebush habitats at JRN were distinct from those at SEV, and pinyonjuniper habitat at BNM was distinct from the pinyon-juniper habitat at SEV (Fig. 6).

Canonical discriminant function analysis (CDFA) tested for differences in ground arthropod taxonomic composition (at the family level) and in abundance among the habitats. The 4 JRN Chihuahuan Desert habitats were significantly different from each other (P < 0.0001; Table 1, Fig. 7a). Standardized canonical coefficient scores revealed that spiders contributed most to the separation of JRN habitats, followed by millipedes, crickets, sun spiders, and bristletails. The Chihuahuan Desert, Rocky Mountain, and Western Short Grassland habitats at SEV were also significantly distinct (P < 0.01; Table 1, Fig. 7b). Beetles contributed most to the separation of SEV habitats, followed by grasshoppers, spiders, centipedes, and millipedes. The Rocky Mountain forest woodland habitats at BNM supported significantly distinct assemblages $(\bar{P} < 0.0001)$, with spiders contributing most to the separation of habitats, followed by bristletails, sun spiders, and crickets (Table 1, Fig. 7c).

Ecoregion Affinities of Selected Ground Arthropods

Two arthropod orders, crickets/grasshoppers (Orthoptera) and spiders (Araneae), were well represented in the pitfall samples by 76 and 121 species, respectively, and were used to examine ecoregion affinities in detail based on published geographic distribution information (Appendix 2). Although many species occurred in ≥ 2 ecoregions, the crickets/grasshoppers and spiders showed different patterns across the sites: orthopteran species richness was most similar between JRN and SEV, while spider species richness was most similar between SEV and BNM. Eight orthopteran species and 16 spider species were found at all 3 sites, representing 10% of the orthopterans and 16% of the spiders (Table 2).

Most of the cricket and grasshopper species (44 species) occurred in the Chihuahuan Desert, followed by lesser numbers of species associated with the Shortgrass Prairie, Southern Rocky Mountains, Colorado Plateau, and

TABLE 1. Canonical discriminant function analysis for differences in ground arthropod taxonomic composition (family level) and abundance among the habitats within each of the 3 study sites. Habitats at Jornada were dominated by mesquite, tarbush, grass, and creosotebush, at Sevilleta by grass, creosotebush, and pinyon/juniper; and at Bandelier by pinyon/juniper, ponderosa pine, and mixed conifers.

	Jornada			Sev	villeta	Bandelier		
Overall multivariate F test (prb. $> F$)	< 0.0001			<	0.01	< 0.0001		
Discriminant variates	1	2	3	1	2	1	2	
Significance (prb. $> F$)	< 0.0001	0.01	0.08	0.01	0.74	< 0.000]	0.01	
Eigenvalue	7.2	2.1	1.2	2.0	0.23	5.9	0.21	
Cumulative variance	68%	88%	100%	89%	100%	90%	100%	

TABLE 2. Total numbers of cricket/grasshopper and spider species found at each and different combinations of our 3 study sites.

	Crickets and grasshoppers	Spiders
Total species sampled	76	121
Jornada only	6	10
Sevilleta only	13	35
Bandelier only	8	29
Jornada and Sevilleta	23	11
Jornada and Bandelier	0	0
Sevilleta and Bandelier	10	15
Jornada, Sevilleta, and Bandelie	er 10	21

Arizona–New Mexico Mountains (Table 3). The Chihuahuan Desert also contributed the greatest number of species unique to an ecoregion (7 species), followed by the Southern Rocky Mountains (6 species). None of the species encountered in this study were unique to the Southern Shortgrass Prairie or to the Colorado Plateau, and only 2 species were unique to the Arizona–New Mexico Mountains.

Spider distributions showed a stronger response to an elevation-precipitation gradient from high richness at high elevation at BNM to lowest richness at JRN. Most spider species were associated with the Arizona–New Mexico Mountains and the Colorado Plateau. Species known from the Chihuahuan Desert and/or the Southern Rocky Mountains also made up a significant part of the fauna. The Shortgrass Prairie region contributed the fewest species. Eight species were reported from only a single ecoregion, and 6 of those were collected from the SEV transition site (Table 3).

DISCUSSION

Local and Regional Community Similarity and Diversity

Results from cluster analysis (at the species level) and CDFA (at the family level) demonstrated that different habitats within a site (JRN, SEV, or BNM) contained distinct ground arthropod assemblages (Figs. 6, 7). Although some species occurred in more than 1 habitat, abundances were not evenly distributed among them. In a similar study, Gering et al. (2003) showed that forest canopy beetles clearly formed different assemblages (with <50% similarity) among the 3 sites in each of 2 ecoregions in forests of Ohio and Indiana. The pattern was maintained for early and late seasons, even with some changes in species composition.

Because of such differences within ecoregions, some researchers may not regard ecoregions as useful delineations for studies of biogeography. Wright et al. (1998) found a poor match between vegetation cover classes and ecoregion boundaries in the American Pacific Northwest. They expected vegetation cover to incorporate information about ecoreigon landforms and climate, but perhaps the varied topography of the region obscured the associations. McDonald et al. (2005) examined species turnover between pairs of adjacent ecoregions and found that similarity declined in the American Southwest, which also is topographically variable. We believe that some groups of plants and animals, and some heterogeneous regions, may not conform well to geographically defined ecoregions or portions of ecoregions. Biogeographic studies involving ecoregions also should address multiple taxa across classes and orders, and examine both species composition and richness.

The importance of regional species source areas to our study was especially clear when we examined arthropod species distributions across the 3 study sites. At this spatial scale, arthropod assemblages that shared common habitats as defined by dominant plant cover (pinyonjuniper habitat at SEV and BNM, desert grassland and creosotebush habitats at SEV and JRN) were not as similar to each other as they

2008]

	Chihuahuan Desert	Shortgrass prairie	Colorado Plateau	Arizona mountains	Southern Rockies
Chihuahuan Desert	64 (3) 44 (7)	44	48	56	42
Shortgrass prairie	27	46 (1) 38 (0)	37	40	34
Colorado Plateau	23	24	77 (1) 34 (0)	67	71
Arizona mountains	21	21	20	78 (2) 29 (2)	57
Southern Rockies	17	22	23	22	65 (1) 36 (6)

TABLE 3. Numbers of cricket/grasshopper species (in bold and below the diagonal) and spider species (above the diagonal) found in this study that are known to be associated with particular ecoregions (based on literature). The numbers in parentheses are species in this study reported to be unique to a particular ecoregion.

were to other habitats within their ecoregions (Fig. 6). In Australia, MacNally et al. (2002) found that the match between ecoregion and faunal diversity depended on the animal taxa studied. Birds and mammals matched ecoregion boundaries more closely than invertebrates and reptiles. MacNally et al. (2002) identified arthropods only to order and acknowledged that greater taxonomic resolution might have improved the strength of regional associations. However, they confirmed that very different taxa do not substitute for each other and that no ecoregion definition will apply to all taxa.

Species Distributions across the Transition Zone

The SEV ecoregion transition site did contain greater species richness and abundance than comparable areas deeper within their respective ecoregions (BNM and JRN, Fig. 4) and the SEV arthropod assemblages were distinct from the other 2 sites (Fig. 6). Evidence for the ecotone or edge effect (Odum 1971) on other ground-dwelling arthropod communities has been mixed. In Israel, Krasnov and Shenbrot (1998) found that the distributions of ground-dwelling darkling beetle (tenebrionid) species did correspond to phytogeographic regions, with increased species diversity at phytogeographic boundaries. In contrast, Bestelmeyer and Wiens (2001) found that the geographic distributions of ground-dwelling ant species did not correspond to phytogeographically defined biomes of the shortgrass steppe (= Western Short Grassland; Ricketts et al. 1999) and Chihuahuan Desert region of North America, nor did ant species diversity increase at biome boundaries. For trees, birds, and

mammals in North America, McDonald et al. (2005) found no marked increase in species turnover rates between pairs of adjacent ecoregions, based on range data and using ecoregion definitions of Ricketts et al. (1999). They did find increased turnover between ecotones that were also on biome boundaries or that followed major mountain ranges. In relation to our study, the SEV site was not only an area of ecoregional transition, it also was in the Southern Rio Grande Rift Valley, which is a transitional area for 3 continental biomes as described in the introduction section. One of our ecoregions, the Southern Rocky Mountains, does track part of a major mountain range, so some of the patterns we observed may be due to these larger geographical features and not just to ecoregions alone.

When we analyzed the ecoregion affinities of crickets and grasshoppers (Orthoptera) and spiders (Araneae) separately, we also found that not all ground arthropods reflected geographic affinities to the same degree. Orthopterans were clearly associated with particular ecoregions, and the overlap in taxa from different ecoregions, particularly Chihuahuan Desert and Western Short Grassland, did contribute to higher species diversity at SEV. Also, as herbivores, grasshoppers tend to specialize on particular plant taxa or groups (Otte and Joern 1977), which themselves tend to be part of the definition of an ecoregion.

Predatory spiders, in contrast, are less closely associated with plant species, although vegetation and substrate structure are important habitat features for them (Wise 1993). Our sites were dominated by the families Gnaphosidae and Lycosidae, both of which have numerous species associated with mesic microhabitats (e.g., Lowrie 1973 for *Pardosa*), as shown by the greatest contribution to our diversity coming from the higher-elevation ecoregions: Colorado Plateau, Arizona-New Mexico Mountains, and Southern Rocky Mountains. The high number of species from the Chihuahuan Desert came in part from families not represented from the higher elevations and from different species within widely distributed families. Our sites also shared 58% of the spider species found in a study in shortgrass steppe in Colorado (Weeks and Holtzer 2000), in the Western Short Grasslands ecoregion, to the north of our sites. The distribution pattern for spiders in our study more closely matches a tracking of the precipitation gradient that Bestelmeyer and Wiens (2001) found in ants. Nevertheless, 6 of the 8 spider species that we sampled that were unique to an ecoregion (as reported in the literature) were from SEV, indicating that biogeographical transitional sites do contribute to regional biodiversity of spiders.

Results from our study did not fully support other likely alternative biogeographic hypotheses-namely, energy or productivity gradients-that would explain our patterns. The energy hypothesis (Hawkins et al. 2003) predicts increasing species diversity with increasing ambient temperature, precipitation, and evapotranspiration, and support for this hypothesis has been found in some arthropod groups (beetles-Kerr and Packer 1999: butterflies-Kerr 2001, Hawkins and Porter 2003). The productivity hypothesis (O'Brien 1998, Whittaker et al. 2003) predicts increasing species richness along with increasing ecosystem productivity, usually measured as plant net primary production (NPP). Temperature and precipitation in the desert Southwest tend to be negatively correlated across elevation gradients (Whittaker and Niering 1975, Brown 1982), often forcing a trade-off between preferred temperature and moisture conditions for arthropods (which are ectothermic animals). Waide et al. (1999) found no consistent relationship between productivity and species diversity for invertebrates within or between communities, or over spatial scales ranging from local (<20 km) to regional (200–4000 km).

Our findings that species composition and richness patterns were consistent with predictions based on ecoregions and edge effects and that these patterns were present among organisms as taxonomically diverse as arthropods in a region as physiographically diverse as the American Southwest indicate that recognition of large-scale biogeographic regions or provinces does have merit. Even though individual species distributions probably are relatively independent from each other, large geographic areas with similar climates and landscapes may support recognizable assemblages of ecologically similar ground arthropod species, as we found. Spatial patterns of terrestrial arthropod communities continue to be poorly understood compared to those of vascular plants and vertebrates. Our findings indicate that consideration of larger geographic source regions, such as ecoregions, may help elucidate patterns of arthropod community species composition and diversity at the local scale.

Acknowledgments

We thank the many field technicians from Jornada LTER, Sevilleta LTER, and Bandelier National Monument who collected arthropod pit-trap samples for this study, especially Dara Parker, Mike Friggens, Karen Wetherill, Kay Beeley, and Rebecca Oertel. We thank George Ball, Jack Brookhart, Richard Fagerlund, Charles O'Brien, Greg Forbes, Paul Johnson, Robert Love, Luis Mendes, Gary Parsons, David Richman, Charles Triplehorn, and David Weissman for arthropod identifications. We thank Teri Neville of the Natural Heritage New Mexico Program for producing the study site map, Paul Neville of Earth Data Analysis Center (EDAC) for providing us with MODIS NPP values, and the data managers from the 3 sites for weather data. Ursula Shepherd and Mike Friggens provided thoughtful comments on earlier versions of the manuscript. This research was funded by National Science Foundation grants to the Jordana (DEB-0080412) and Sevilleta (DEB-0080529) LTER programs, and research at Bandelier National Monument was funded by the National Park Service and U.S. Geological Survey (BRD Global Change Research Program, Western Mountain Initiative).

LITERATURE CITED

ADIS, J. 1979. Problems of interpreting arthropod sampling with pitfall traps. Zoologische Anzeiger 202:177–184.

ARNETT, R.H., JR. 2000. American insects. CRC Press, Boca Raton, FL. 1003 pp.

- AXLEROD, D.I. 1958. Evolution of the Madro-Tertiary geoflora. Botanical Review 24:433–509.
- BESTELMEYER, B.T., AND J.A. WIENS. 2001. Local and regional-scale responses of ant diversity to a semiarid biome transition. Ecography 24:381–392.
- BROWN, D.E., EDITOR. 1982. Biotic communities of the American Southwest: United States and Mexico. Desert Plants 4:1–342.
- BROWN, J.H. 1995. Macroecology, University of Chicago Press, Chicago, IL. 269 pp.
- BROWN, J.H., AND M.V. LOMOLINO. 1998. Biogeography. Sinauer and Associates, Sunderland, MA. 691 pp.
- CRAWFORD, C.S. 1981. Biology of desert invertebrates. Springer-Verlag, Berlin. 314 pp.
- DICK-PEDDIE, W.A. 1993. New Mexico vegetation: past, present, and future. University of New Mexico Press, Albuquerque. 244 pp.
- DIGWEED, S.C., C.R. CURRIE, H.A. CARCAMO, AND J.R. SPENCE. 1995. Digging out the "digging-in effect" of pitfall traps: influences of depletion and disturbance on catch of ground beetles (Coleoptera: Carabidae). Pedobiologia 39:561–576.
- GERING, J.C., T.O. CRIST, AND J.A. VEECH. 2003. Additive partitioning of species diversity across multiple spatial scales: implication for regional conservation of biodiversity. Conservation Biology 17:488–499.
- HAWKINS, B.A., R. FIELD, H.V. CORNELL, D.J. CURRIE, J.F. GUEGAN, D.M. KAUFMAN, J.T. KERR, ET AL. 2003. Energy, water, and broad-scale geographic patterns of species richness. Ecology 84:3105–3117.
- HAWKINS, B.A., AND E.E. PORTER. 2003. Water-energy balance and the geographic pattern of species richness of western Palearctic butterflies. Ecological Entomology 28:678–686.
- HAWLEY, J.W. 1978. Guidebook to the Rio Grande Rift in New Mexico and Colorado. New Mexico Bureau of Mines and Mineral Resources Circular 163:1–124.
- HOLLAND, M.M., P.G. RISSER, AND R.J. NAIMAN. 1991. Ecotones: the role of landscape boundaries in the management and restoration of changing environments. Chapman and Hall, NY. 142 pp.
- HUNT, C.B. 1983. Physiographic overview of our arid lands in the western U.S. Pages 7–63 *in* S.G. Wells and D.R. Haragan, editors, Origin and evolution of deserts. University of New Mexico Press, Albuquerque.
- KERR, J.T. 2001. Butterfly species richness patterns in Canada: energy, heterogeneity, and the potential consequences of climatic change [online]. Conservation Ecology 5:10. Available from: http://www.consecol .org/vol5.iss/art10
- KERR, J.T., AND L. PACKER. 1999. The environmental basis of North American species richness patterns among Epicauta (Coleoptera: Meloidae). Biodiversity and Conservation 8:617–628.
- KRASNOV, B.R., AND G.I. SHENBROT. 1998. Structure of communities of ground-dwelling animals at the junction of two phytogeographic zones. Journal of Biogeography 25:1115–1131.
- LOMOLINO, M.V., AND L.R. HEANEY, EDITORS. 2004. Frontiers of biogeography: new directions in the geography of nature. Sinauer and Associates, Sunderland, MA. 436 pp.
- LOWRIE, D.C. 1973. The microhabitats of western wolf spiders of the genus *Pardosa*. Entomological News 84:103–116.
- MACNALLY, R., A.F. BENNETT, G.W. BROWN, L.F. LUMSDEN, A. YEN, S. HINKLEY, P. LILLYWHITE, AND D. WARD.

2002. How well do different ecosystem-based planning units represent different components of biodiversity? Ecological Applications 12:900–912.

- MCDONALD, R., M. MCKNIGHT, D. WEISS, E. SELIG, M. O'CONNOR, C. VIOLIN, AND A. MOODY. 2005. Species compositional similarity and ecoregions: do ecoregion boundaries represent zones of high species turnover? Biological Conservation 126:24–40.
- O'BRIEN, E.M. 1998. Water-energy dynamics, climate, and prediction of woody plant species richness: an interim general model. Journal of Biogeography 25:379–398.
- ODUM, E.P. 1971. Fundamentals of ecology. 3rd edition. W.B. Saunders Co., Philadelphia, PA. 574 pp.
- OTTE, D., AND A. JOERN. 1977. On feeding patterns in desert grasshoppers and the evolution of specialized diets. Proceedings of the Academy of Natural Sciences of Philadelphia 128:89–126.
- PARMENTER, R.R., S.L. BRANTLEY, J.H. BROWN, C.S. CRAWFORD, D.C. LIGHTFOOT, AND T.L. YATES. 1995. Diversity of animal communities on Southwestern rangelands: species patterns, habitat relationships, and land management. Pages 50–71 in N. West, editor, Biodiversity on rangelands: natural resources and environmental issues. Volume 4. College of Natural Resources, Utah State University, Logan.
- POLIS, G.A., EDITOR. 1991. The ecology of desert communities. University of Arizona Press, Tucson. 456 pp.
- POOLE, R.W., AND P. GENTILI, EDITORS. 2003. Nomina Insecta Nearctica. Entomological Information Services, Rockville, MD. 4 volumes, 3494 pp.
- RICKETTS, T.H., E. DINERSTEIN, D.M. OLSON, C.J. LOUCKS, W. EICHBAUM, D. DELLASALA, K. KAVANAGH, ET AL. 1999. Terrestrial ecoregions of North America: a conservation assessment. World Wildlife Fund, Island Press, Washington, DC. 485 pp.
- RICKLEFS, R.E., AND G.L. MILLER. 2000. Ecology. 4th edition. W.H. Freeman and Co., New York. 822 pp.
- RICKLEFS, R.E., AND D. SCHLUTER, EDITORS. 1993. Species diversity in ecological communities: historical and geographical perspectives. University of Chicago Press, Chicago, IL. 414 pp.
- RISSER, P.G. 1993. Ecotones at local to regional scales from around the world. Ecological Applications 3: 367–368.
- _____. 1995. The status of the science examining ecotones. BioScience 45:318–325.
- SAS INSTITUTE, INC. 2003. PC SAS, version 8.2. Statistical Analysis Systems, SAS Institute, Inc., Cary, NC.
- SHMIDA, A., AND M.W. WILSON. 1985. Biological determinants of species diversity. Journal of Biogeography 12:1–20.
- SOUTHWOOD, T.R.E. 1966. Ecological methods: with particular reference to the study of insect populations. 2nd edition. Halsted Press, John Wiley & Sons, Inc., New York. 391 pp.
- THOMAS, D.B., JR., AND E.L. SLEEPER. 1977. The use of pit-fall traps for estimating the abundance of arthropods, with special reference to the Tenebrionidae (Coleoptera). Annals of the Entomological Society of America 70:242–248.
- UETZ, G.W., AND J.D. UNZICKER. 1976. Pitfall trapping in ecological studies of wandering spiders. Journal of Arachnology 3:101–111.
- VAN DEVENDER, T.R. 1995. Desert grassland history: changing climates, evolution, biogeography, and

Downloaded From: https://bioone.org/journals/Western-North-American-Naturalist on 08 Jan 2020 Terms of Use: https://bioone.org/terms-of-use Access provided by United States Department of Agriculture National Agricultural Library (NAL) community dynamics. Pages 68–99 *in* M.P. McClaren and T.R. Van Devender, editors, The desert grassland. University of Arizona Press, Tucson.

- VAN DEVENDER, T.R., AND W.G. SPAULDING. 1983. Development of vegetation and climate in the southwestern United States. Pages 131–156 in S.G. Wells and D.R. Haragan, editors, Origin and evolution of deserts. University of New Mexico Press, Albuquerque.
- WAIDE, R.B., M.R. WILLIG, C.F. STEINER, G. MITTEL-BACH, L. GOUGH, S.I. DODSON, G.P. JUDAY, AND R. PARMENTER. 1999. The relationship between productivity and species richness. Annual Review of Ecology and Systematics 30:257–300.
- WARD, D.F., T.R. NEW, AND A.L. YEN. 2001. Effects of pitfall trap spacing on the abundance, richness and composition of invertebrate catches. Journal of Insect Conservation 5:47–53.
- WEEKS, R.D., JR., AND T.O. HOLTZER. 2000. Habitat and season in structuring ground-dwelling spider (Araneae) communities in a shortgrass steppe ecosystem. Environmental Entomology 29:1164–1172.
- WHITFORD, W.G. 1986. Decomposition and nutrient cycling in deserts. Pages 93–117 in W.G. Whitford, editor, Pattern and process in desert ecosystems. University of New Mexico Press, Albuquerque.
- WHITTAKER, R.H. 1975. Communities and ecosystems. 2nd edition. Macmillan Publishing Company, New York. 385 pp.

- WHITTAKER, R.H., AND W.A. NIERING. 1975. Vegetation of the Santa Catalina Mountains, Arizona, USA. Part 5, Biomass production and diversity along the elevation gradient. Ecology 56:771–790.
- WHITTAKER, R.J., K.J. WILLIS, AND R. FIELD. 2003. Climatic-energetic explanations of diversity: a macroscopic perspective. Pages 107–129 in T.L. Blackburn and K.J. Gaston, editors, Macroecology: concepts and consequences. Blackwell Science, Oxford, U.K.
- WISE, D.H. 1993. Spiders in ecological webs. Cambridge University Press, Cambridge, U.K. 328 pp.
- WOODCOCK, B.A. 2005. Pitfall trapping in ecological studies. Pages 37–57 in S.R. Leather, editor, Insect sampling in forest ecosystems. Blackwell Publishing, Oxford, U.K.
- WRIGHT, R.G., M.P. MURRAY, AND T. MERRILL. 1998. Ecoregions as a level of ecological analysis. Biological Conservation 86:207–213.
- ZHAO, M., F.A. HEINSCH, R.M. RAMAKIRSHNA, AND S.W. RUNNING. 2005. Improvements of the MODIS terrestrial gross and net primary production global data set. Remote Sensing of Environment 95:164–176.
- ZHAO, M., AND S.W. RUNNING. 2006. Sensitivity of moderate resolution imaging spectroradiometer (MODIS) primary production to the accuracy of meterological reanalyses. Journal of Geophysical Research 111:1–13.

Received 22 January 2007 Accepted 10 October 2007 APPENDIX 1. List of target arthropod species included in this study and their occurrence by site. BNM = Bandelie E T

in this study and their occurrence blier National Monument, SEV =				Taxon	BNM	SEV	JR
Ecological Research areas, JRN = Term Ecological Research areas.	= Jornada	Basin	Long-	Dictyna sp. 1			
				Dictyna sp. 2		11	
Taxon	BNM	SEV	JRN	dictynid immatures Family Filistatidae		4	
Class Chilopoda – centipedes				Filistatinella sp.		14	
Order Geophilomorpha				Kukulcania hibernalis		3	
Family Diagnothodontidae				Family Gnaphosidae		5	
Strigamia chionophila	16			Callilepis chisos		3	
Order Lithobiomorpha	10			C. mumai	6	16	
Family Lithobiidae				Cesonia sincera	0	36	
Anobius centurio	1			Drassodes gosiutus	2	30	2
Lithobius forficatus			4	Dd. neglectus	6	1	_
Nadabius holzingeri	9			Dd. saccatus	2	53	
Nadabius mesenchinus	47			Drassyllus antonito		1	
Nadabius pullus	12			Dr. conformans		8	
Paitobius juventus	2			Dr. dromeus	8	34	
Pokabius bilabiatus	3			Dr. insularis group		1	
Taiubius harrietae	2	2		Dr. lamprus		3	
Order Scolopendromorpha				Dr. lepidus		1	
Family Scolopendridae				Dr. mexicanus		6	
Scolopendra polymorpha	1	53	3	Dr. mormonus		7	
Class Diplopoda – millipedes				Dr. mumai		42	
Order Parajulida				Dr. orgilis	1	2	
Family Parajulidae				Gnaphosa clara	1	2	
Oriulus medianus	259			G. muscorum	92		
parajulid sp.		14		Halpodrassus bicornis	2		
Order Polydesmida				Ha. chamberlini	14	133	2
Family Polydesmidae				Ha. dixianus	8		
Utadesmus hoffi	2			Ha. eunis	3		
Order Spirostreptida				Ha. signifer	17		
Family Spirostreptidae		-	0	Herpyllus bubulcus	1	25	2
Orthoporus ornatus		7	8	He. cockerelli	5		
Class Arachnida				He. excelsus		1	
Order Araneae – spiders				He. hesperolus	1	34	
Family Agelenidae			1	He. propinquus		2	
Agelenopsis sp. A. longistylus		12	1	Micaria aenea	3		
A. tongistytus Agelenopsis utahensis	2	2		M. gosiuta		4	
Hololena hola	39	17		M. imperiosa		10	
Novalena lutzi	5	11		M. longipes		,	1
agelenid immatures	6	33		M. porta	4	1	
Family Anyphaenidae	0	00		M. pulicaria	4		
Anyphaena dixiana		16		M. nr. triangulosa		1	
A. hespar	13			Nodocion utus Orodrassus coloradensis	8	1	
A. marginalis	2			Zelotes anglo	2	24	
A. pacifica	2			Zeibles angio Z. fratris	23	24	
Hibana incursa		3		Z. lasalanus	23 7	22	
anyphaenid immatures	26	38		Z. puritanus Z. puritanus	12	22	
Family Clubionidae				Z. tuobus	21	80	1
Clubiona sp.		53		gnaphosid immatures	90	1599	1
clubionid immatures		25		Family Linyphiidae	00	1000	
Family Corinnidae				combined species	88	107	
Castianeira sp. 1	3			Pityohyphantes sp.	14	101	
Castianeira sp. 2			19	Family Lycosidae	11		
Castianeria sp. 3		3		Alopecosa kochi	130	112	
C. occidentalis		71		Geolycosa rafaelana	100	4	
Meriola decepta		2		Hogna carolinensis	1	35	
corinnid immatures		3		H. frondicola	6	2	
Family Dictynidae				Pardosa montanensis	23	_	
Cicurina deserticola	1			P. orophila	5	177	
C. robustus	31		_	P. uncata	503		
Cicurina spp.	72	98	9	P. xerophila	12		
C. varians	11		3	1. легорини			

Downloaded From: https://bioone.org/journals/Western-North-American-Naturalist on 08 Jan 2020 Terms of Use: https://bioone.org/terms-of-use Access provided by United States Department of Agriculture National Agricultural Library (NAL)

2008]

Appendix	1.	Continued
----------	----	-----------

Appendix 1. C	Continued.
---------------	------------

ion	BNM	SEV	JRN	Taxon	BNM	SEV	JR
Schizocosa chiricahua		12		Eurybunus brunneus	6		
S. mccooki	60	39	8	sclerosomatid sp.	0	15	
S. mimula	2	1		Togwoteeus biceps	271		
Trochosa terricola	11	2		Trachyrhinus marmoratus	624		
Varacosa gosiuta	34	17	2	Family Triaenonychidae			
lycosid immatures	193		2	Sclerobunus robustus	14		
Family Liocranidae				Order Scorpiones – scorpions			
Neoanagraphis chamberlini		3	1	Family Vaejovidae			
Family Mimetidae				Vaejovis coahuilae		523	13
Mimetus hesperus		8	2	V. russelli		2	
Family Miturgidae			2	Order Uropygi – vinegaroons			
Cheiracanthium inclusum Syspira sp.			17^{2}	Family Thelyphonidae Mastigoproctus giganteus		14	
Family Oxyopidae			17	Class Insecta		14	
Oxyopes lynx		6		Order Archaeognatha – bristletail	s		
O. salticus		2		Family Meinertellidae	5		
O. tridens		-	27	Machilinus aurantiacus	788	1199	
Family Philodromidae				Praemachilellus rentzi			5^{4}
Apollophanes texanus		10	31	Family Machilidae			
Ebo sp.	3			Mesomachilis sp.		3	
Ebo parabolis		22	24	Order Dictyoptera – cockroaches,	mantise	s	
Philodromus infuscatus	1			Family Blattellidae			
P. keyserlingi		3		Parcoblatta desertae			2
Thanatus coloradensis	1	51	10	Family Polyphagidae			
Tibellus duttoni		8		Arenivaga erratica		54	(
Family Salticidae		~~		Eremoblatta subdiaphana		6	
Habronattus clypeatus	1	55		Family Mantidae		C	
H. conjunctus	1	1 9	14	Litaneutria minor		6 3	
H. geronimoi Metacyrba taeniola		9	14 2	<i>Yersiniops solitaria</i> Order Orthoptera – grasshoppers,	kotydid		ote
Pellenes limatus		1	4	Family Acrididae	катуши	s, crick	ets
Phidippus spp.	5	6	5	Acantherus piperatus		35	Ę
Platycryptus arizonenesis	0	14	0	Aeoloplides elegans		00	
Sassacus sp.			12	Ageneotettix deorum		48	
salticid immatures	8	110		Amphitornus coloradus		16	
Family Sicariidae				Arphia conspersa	22	410	
Loxosceles apachea		4	2	A. pseudonietana	5	54	
Family Theridiidae				Aulocara femoratum		444	
Euryopes scriptipes	17	8	6	Barytettix humphreysi		9	
Latrodectus hesperus	22	35	8	Bootettix argentatus		280	2
<i>Steatoda</i> sp.	-	1		Camnula pellucida	1		
S. fulva	2	7	11	Campylacantha olivacea	10		-
S. hespera	5		,	Chorthippus curtipennis	16	00	
theridiid immatures Family Thomisidae	7		1	Cibolacris parviceps		98	4
Bassaniana versicolor		2		Clematodes larreae Cordillacris crenulata		1	
Misumenops sp.		1		C. occipitalis		23	
Xysticus apachecus		9		Dactylotum bicolor		29	
X. cunctator	6	6	1	Eritettix simplex	6	57	
X. facetus	1	24	3	Heliaula rufa		19	
X. gosiutus	1			Hesperotettix viridis	1	2	
X. gulosus	2	10		Hippopedon capito		3	
X. lassanus	1	77	9	L. wheeleri		6	
X. locuples	6	11		Ligurotettix planum			
X. luctuosus	9			Melanoplus aridus	6	2	
X. montanensis	19			M. arizonae		53	
X. orizaba		7		M. bowditchi		56	
thomisid immatures	13	168	22	M. femur-nigrum	3		
Order Solifugae – solifugids, sun s	piders			M. gladstoni	3	26	
Family Eremobatidae	000	250	100	M. lakinus	0	10	
Eremobates sp.	232	253	129	M. occidentalis	3		
Order Opiliones – harvestmen				M. packardi		1	

WESTERN NORTH AMERICAN NATURALIST

APPENDIX 1. Continued.

APPENDIX 1. Continued.					APPENDIX 1. Continued.				
laxon		BNM	SEV	JRN	Taxon	BNM	SEV	JRI	
Me	estobregma plattei		21		Lygaeus kalmi		43	ļ	
	peia obscura		18	3	Melacoryphus lateralis			5	
	ropomala pallida		15	4	Ozophora sp.		172		
Ph	lebostroma quadrimaculati	um	3		Prytanes sp.		2		
	oetaliotes nebracensis		2		Sphragisticus sp.		1		
Pse	oloessa delicatula		227	3	Family Miridae				
<i>P.</i> 1	texana	37	700	8	Eustictus sp.		560		
Sp	haragemon equale		2		Order Coleoptera – beetles				
	rbula admirabilis		7		Family Anobiidae				
	montezuma		53		anobiid sp.			2	
	achyrhachys kiowa		9	1	Niptus ventriculus				
	imerotropis californica		8	4	Family Carabidae				
	cincta	6	2	-	Amara sp.	1			
	modesta	3	-		A. apachensis		5		
	pallidipennis	67	940	86	A. discors		8		
	pistrinaria	01	12	00	A. erratica		4		
	suffusa	16	12		A. nr. idahoana		20		
	opidolophus formosus	10	9	5	A. rubrica		30		
	inthippus corallipes	4	54	6	Calosoma obsoletus		2		
	ridid immatures	55	642	19	C. peregrinator		12		
	ly Gryllidae	55	042	15	Carabus taedatus	787			
	<i>yllus</i> sp. 1 undescribed		428	613	Cicindela lemniscata	101	4		
				015	C. pulchra		13		
	ryllus pennsylvanicus		314		C. punctulata		28		
	personatus	110	439		Cymindis arizonensis	1	49		
	<i>yllus</i> sp. 2 undescribed	113	10		C. cribricollis	2	49		
	ecanthus californicus	14	12	~		9	110	1	
	yllid immatures		733	5	C. punctigera	9 2	110	1	
	ly Mogoplistidae	20	2024	07	Dyschirius sp.	Z	F		
	cloptilium comprehendens	26	2634	37	D. globulosa		5		
	plosphyrum boreale		363		Euryderus grossus		10		
	ly Rhaphidophoridae				Harpalus amputatus		12		
	nmobaenetes phrixocnemoi	des	_	30	H. katiae		1		
	euthophilus lamellipes		5		H. pennsylvanicus		1		
	pallidus	195	155		H. tadorcus		149		
	utahensis	4111	38		Helluomorphoides ferruginea		1		
	variegatus			53	He. latitarsus		1		
	uhiniodes hastiferum		919	7	Lebia viridis	2	21		
Stų	yracosceles neomexicana	3			Notiophilus novemstriatus	19			
	aphidophorid immatures		922		Pasimachus californicus		56	5	
Fami	ly Stenopelmatidae				P. elongatus/obsoletus		1693]	
Ste	enopelmatus fuscus	153	130		Poecilus lucublandus		1		
	enopelmatus sp. 1 undescri	bed	9		Pterostichus adstrictus	1019			
Fami	ly Tettigoniidae				Rhadine dissectus		2		
Ar	ethaea gracilipes	2	33	9	Rhadine sp. 3		1		
	pnobotes fuliginosus		1	1	R. umbra group	24			
	chopetala brevihastata			5	Scaphinotus snowi	2			
	emopedes balli	1	27		Scarites subterraneus		5		
	bilineata		14		Tachys sp.	1			
	scudderi		237	34	Family Cryptophagidae				
	sara elegans		9	37	cryptophagid sp. 1	14	116		
	sara juniperi	3	7	2.	Cryptophagus dentatus	19]	
	ttigoniid immatures	0	41	1	C. fumidulus	10	1		
	Iemiptera – true bugs			-	C. stromus		9		
	ly Aradidae				Myrmedophila americanus	1	0		
	adus lugubris	9			Family Curculionidae	T			
	ly Cydnidae	9			Apion sp.		1		
			1894		Apion sp. Cimbocera buchanani		31		
	dnid sp.	C	1884	C				1	
	ıllasiellus discrepans	6	1197	6	C. conspersa		2]	
	ly Lygaeidae		010	10	Cleonidius poricollis		1		
	nblethis vicarius	33	310	40	Crocidema sp.	_	4		
	eocoris sp.		5		curculionid sp.	1			
lve	gaeid sp. 2	1			Gerstaeckeria lecontei		17		
	gaeid sp. 28		2		G. turbida		1		

APPENDIX 1. Continued.

2008]

BIOGEOGRAPHY OF GROUND-DWELLING ARTHROPODS

APPENDIX 1. Continued.

						-	
ion	BNM	SEV	JRN	Taxon	BNM	SEV	JRN
Hypera postica		1		Anatropis verticalis			152
Minyomerus languidus		8	3	Aphodius sp. 1		4	
Notiodes limatulus		8		Aphodius sp. 2			
Ophryastes sp.			7	Aphodius sp. 3		1	
O. dunnianus		1		Boreocanthon ebenus			
O. globularis		45		B. probus		1	
O. latirostris		1		Canthon puncticollis			
O. sulcatus		3	3	Cremastocheilus planatus		2	
O. vittatus		1	2	Diplotaxis sp. 1	17		
Pandeleteius dentipes		3		Diplotaxis sp. 2		32	
Rhypodillus brevicollis		1		Diplotaxis sp. 3		114	
Sapotes longipilis		37		D. carbonata	1		1
S. puncticollis		5		D. knausi			1
Scyphophorus acupunctatus		2		D. subangulata	6	25	
Sitona hispidulus		1		Euphoria inda	127	37	
Smicronyx sp.			1	Hoplia laticollis		1	
Sphenophorus sp.	1			Onthophagus sp.		1	
Yuccaborus frontalis		5		O. hecate			
Family Elateridae				Paracotalpa puncticollis		13	
Aeolus mellillus		2		Phyllophaga sp.	5		
Agrypnus rectangularis	50	-	6	P. vetula	3	14	
Cardiophorus sp.	00	89	0	P. wickhami	0	160	
Conoderus athoides		27	5	scarabaeid sp. 46		16	
Ctenicara carbo		4	5	scarabaeid sp. 51		10	
C. pudica	3	-1		scarabaeid sp. 57		4	
Glyphonyx sp.	1			scarabaeid sp. 60		1	
Heteroderes sordidus	1	2				1	
Horistonotus simplex		2 19		Family Staphylinidae	1265	G	
	0		101	aleocharine spp.		6	
Lanelater schotti	9	1816	121	Carphacis nepigonensis	4		
Limonius lanei	7			Dexiogyia sp.	11		
Melanotus similis		1	_	Hapalaraea cacti	10		
Neotrichophorus arizonensis		2	7	Lordithon arizonensis	46		
Family Endomychidae				omaliine sp. 1	1	11	
Aphorista sp.	1			Ocypus ater		3	
Family Erotylidae				Oxytelus sp.	2		
Cypherotylus californicus	3			Parothius sp.	2		
Family Histeridae				Philolonthus sp.	1		
Hister abbreviatus	9		32	Pseudopsis callosa	3		
Iliotona cacti		5	1	Quedius desertus	1	4	
Saprinus sp.	1			staphylinine sp. 1	1		
S. discoidalis		34		staphylinine sp. 2	1		
S. pennsylvanicus		5		Tachyporus jocusus	55		
Xerosaprinus sp. 1		0	1	Family Tenebrionidae	00		
Xerosaprinus sp. 2		399	-	Anepsius sp.		221	
Family Lathridiidae		000		Araeoschizus decipiens		194	3
Corticaria rudis	8			Argoporis rufipes		50	C.
Family Leiodidae	0			Blapstinus fortis	1	1	
	1			B. pimalis	1	61	
Catops basilarus	1	1	10			01	1
Ptomaphagus texanus	14	1	16	Conibius uniformis		~	1
Family Nitidulidae	2			Edrotes leechi		5	
Carpophilus sp.	2			Ed. rotundus		23	
C. humeralis	67	-		Eleodes carbonarius		6	
C. lugubris	45	2	69	E. caudiferus		48	
C. palllidipennis	26			E. debilis	1		
C. sayi			3	E. extricatus	594	1193	
Epuraea adumbrata	4			E. fusiformis		9	
Phenolia grossa	15			E. gracilis		369	
Thalycra keltoni	7			E. hispilabris		15	12
Family Ptiliidae				E. hoppingi	80		
Acrotrichus sp.	83			E. longicollis	8	1410	3
Family Salpingidae	00			E. nigrinus	99		0
Elacatis umbrosus	2			E. obscurus	339		
	-				550	09	
Family Scarabaeidae				E. obsoletus		92	

APPENDIX 1. Continued.

APPENDIX 1. Continued.				APPENDIX 1. Continued.					
Taxon	BNM	SEV	JRN	Taxon	BNM	SEV	JRN		
E. pimelioides	8			Me. coarcticollis		3			
E. sponsus		33		Metopoloba pruinosus		9			
E. tenuipes		94		Metaponium sp. 1	12				
E. tricostatus		17		Mt. cribriceps	1	1	183		
Embaphion contusum	31	1	1	Mt. implicans	1				
Em. glabrum			3	Neobaphion planipennis		24			
Em. planum		10		Stenomorpha consors		5			
Eupsophulus castaneus			3	S. convexicollis		16	9		
Eusattus reticulatus		3	3	S. obovata		29			
Glyptasida sordida		16	2	S. rimata		1			
Gonasida elata		6		S. severa		7			
Helops callosa		8		Steriphanus convexus	30	540			
Lobometopon fusiformis		12		Telabis histricus		1			
Megasida obliterata		32		Trimytis pruinosa		28	22		
M. tenuicollis		173	32	Trogloderis costatus		3			
Melanastus sp. 1		345							

ADDET JDIV 1 Continued

APPENDIX 1 Continued

APPENDIX 2. References for the geographic distributions of spider, cricket, and grasshopper species relative to the Southern Rio Grande Rift.

- BRADY, A.R. 1964. The lynx spiders of North America, north of Mexico (Araneae: Oxyopidae). Bulletin of the Museum of Comparative Zoology 131:429–518. . 1979. Nearctic species of the wolf spider genus
- Trochosa (Araneae: Lycosidae). Psyche 86:167–212. CHAMBERLIN, R.V. 1923. The North American species of
- Mimetus. Journal of Entomology and Zoology (Claremont) 15:3–9.
- . 1924. The spider fauna of the shores and islands of the Gulf of California. Proceedings of the California Academy of Sciences 12:561–694.
- CHAMBERLIN, R.V., AND W. IVIE. 1940. Agelenid spiders of the genus *Cicurina*. Bulletin of the University of Utah 30:1–107.
- _____. 1941. North American Agelenidae of the genera Agelenopsis, Calilena, Ritalena and Tortolena. Annals of the Entomological Society of America 34:585–628.
- _____. 1942. Agelenidae of the genera Hololena, Novalena, Rualina and Melpomene. Annals of the Entomological Society of America 35:203–241.
- DONDALE, C.D., AND J.H. REDNER. 1969. The *infuscatus* and *dispar* groups of the spider genus *Philodromus* in North and Central America and the West Indies (Araneida: Thomisidae) Canadian Entomologist 101:921–954.
 - . 1975. Revision of the spider genus *Apollophanes* (Araneida: Thomisidae). Canadian Entomologist 107: 1175–1192.
- 1976. A review of the spider genus *Philodromus* in the Americas (Araneida: Philodromidae). Canadian Entomologist 108:127–157.
 - . 1978. Revision of the nearctic wolf spider genus Schizocosa (Araneae: Lycosidae). Canadian Entomologist 110:143–181.
- _____. 1979. Revision of the wolf spider genus *Alopecosa* Simon in North America (Araneae: Lycosidae). Canadian Entomologist 111:1033–1055.
- _____. 1990. The wolf spiders, nurseryweb spiders and lynx spiders of Canada and Alaska: Araneae: Lycosidae, Pisauridae, and Oxyopidae. Insects and Arachnids of Canada 17:1–383.
- DONDALE, C.D., A.L. TURNBULL, AND J.H. REDNER. 1964. Revision of the Nearctic species of *Thanatus* C.L. Koch (Araneae: Thomisidae). Canadian Entomologist 96:636–656.
- EDWARDS, R.J. 1958. The spider subfamily Clubioninae of the United States, Canada and Alaska (Araneae: Clubionidae). Bulletin of the Museum of Comparative Zoology 118:365–436.
- GERTSCH, W.J. 1933. Notes on American spiders of the family Thomisidae. American Museum Novitates 593:1–22.
- _____. 1935. Spiders from the southwestern United States, with descriptions of new species. American Museum Novitates 792:1–31.
- . 1939. A revision of the typical crab-spiders (Misumeninae) of America north of Mexico. Bulletin of the American Museum of Natural History 76:277–442.
- _____. 1953. The spider genera *Xysticus, Coriarachne*, and *Oxyptila* (Thomisidae, Misumeninae) in North

APPENDIX 2. Continued.

America. Bulletin of the American Museum of Natural History 102:417–482.

- GERTSCH, W.J., AND F ENNIK. 1983. The spider genus Loxosceles in North America, Central America, and the West Indies. Bulletin of the American Museum of Natural History 175:264–360.
- HELFER, J.R. 1987. How to know the grasshoppers, crickets, cockroaches, and their allies. Dover Publications, Inc., New York. 363 pp.
- HUBBELL, T.H. 1936. A monographic revision of the genus *Ceuthophilus* (Orthoptera, Gryllacrididae, Raphidophorinae). University of Florida Biological Series, 2, 1–551.
- KASTON, B.J. 1970. Comparative biology of American black widow spiders. Transactions of the San Diego Society of Natural History 16:33–82.
- LEVI, H.W. 1954. Spiders of the genus *Euryopis* from North and Central America (Araneae, Theridiidae). American Museum Novitates 1666:1–48.
- _____. 1957. The spider genera Crustulina and Steatoda in North America, Central America, and the West Indies (Araneae, Theridiidae). Bulletin of the Museum of Comparative Zoology 117:367–424.
- LOWRIE, D.C., AND C.D. DONDALE. 1981. A revision of the *nigra* group of the genus *Pardosa* in North America. Bulletin of the American Museum of Natural History 170:125–139.
- OTTE, D. 1981. The North American grasshoppers. Volume 1. Acrididae, Gomphocerinae and Acridinae. Harvard University Press, Cambridge, MA. 275 pp.
- . 1984. The North American grasshoppers. Volume 2. Acrididae, Oedopodinae. Harvard University Press, Cambridge, MA. 376 pp.
- PLATNICK, N.I. 1974. The spider family Anyphaenidae in America north of Mexico. Bulletin of the Museum of Comparative Zoology 146:205–266.
- _____. 1975. A revision of the holarctic spider genus *Callilepis* (Araneae, Gnaphosidae). American Museum Novitates 2573: 1–32.
- PLATNICK, N.I., AND A. LAU. 1975. A revision of the *celer* group of the spider genus *Anyphaena* (Araneae, Anyphaenidae) in Mexico and Central America. American Museum Novitates 2575:1–36.
- PLATNICK, N.I., AND M.U. SHADAB. 1974. A revision of the bispinosus and bicolor groups of the spider genus Trachelas (Araneae, Clubionidae) in North and Central America and the West Indies. American Museum Novitates 2560:1–34.
- _____. 1975a. A revision of the spider genus *Gnaphosa* (Araneae, Gnaphosidae) in America. Bulletin of the American Museum of Natural History 155:1–65.
- _____. 1975b. A revision of the spider genera *Haplodra*sus and *Orodrassus* (Araneae, Gnaphosidae). American Museum Novitates 2583:1–40.
- _____. 1976. A revision of the spider genera Drassodes and Tivodrassus (Araneae, Gnaphosidae) in North America. American Museum Novitates 2593:1–29.
- ______. 1977. A revision of the spider genera *Herpyllus* and *Scotophaeus* (Araneae, Gnaphosidae) in North America. Bulletin of the American Museum of Natural History 159:1–44.
- . 1980a. A revision of the North American spider genera Nodocion, Litopyllus, and Synaphosus (Araneae,

APPENDIX 2. Continued.

APPENDIX 2. Continued.

- Gnaphosidae). American Museum Novitates 2691: 1–26.
- _____. 1980b. A revision of the spider genus Cesonia (Araneae, Gnaphosidae). Bulletin of the American Museum of Natural History 165:335–386.
 - ____. 1982. A revision of the American spiders of the genus *Drassyllus* (Araneae, Gnaphosidae). Bulletin of the American Museum of Natural History 173:1–97.
- _____. 1983. A revision of the American spiders of the genus *Zelotes* (Araneae, Gnaphosidae). Bulletin of the American Museum of Natural History 174:97–192.
- _____. 1988. A revision of the American spiders of the genus *Micaria* (Araneae, Gnaphosidae). American Museum Novitates 2916:1–64.
- RICHMAN, D.B., D.C. LIGHTFOOT, C.A. SUTHERLAND, AND D.J. FERGUSON. 1993. A manual of the grasshoppers of New Mexico. Handbook No. 7, Cooperative Extension Service, New Mexico State University, Las Cruces. 112 pp.

- SAUER, R.J., AND N.I. PLATNICK. 1972. The crab spider genus *Ebo* (Araneida: Thomisidae) in the United States and Canada. Canadian Entomologist 104:35–60.
- VETTER, R.S. 2001. Revision of the spider genus Neoanagraphis (Araneae, Liocranidae). Journal of Arachnology 29:1–10.
- VOGEL, B.R. 1964. A taxonomic revision of the distincta group of the wolf-spider genus Pardosa in America north of Mexico (Araneida, Lycosidae). Postilla 82:1–30.
- . 2004. A review of the spider genera *Pardosa* and *Acantholycosa* (Araneae, Lycosidae) of the 48 contiguous United States. Journal of Arachnology 32:55–108.
- WALLACE, H.K. 1942. A revision of the burrowing spiders of the genus *Geolycosa* (Araneae, Lycosidae). American Midland Naturalist 27:1–62.